

DOI: 10.21767/2386-5180.100260

Resistance to Tetracycline and Vancomycin of *Staphylococcus aureus* Isolates from Sanandaj Patients by Molecular Genotyping

Foozieh Arabzadeh¹, Fatemeh Aeini², Fatemeh Keshavarzi^{2*} and Sahra Behrvash²¹Department of Biology, Kurdistan Science and Research Branch, Islamic Azad University, Sanandaj, Iran²Department of Biology, Sanandaj Branch, Islamic Azad University, Sanandaj, Iran

*Corresponding author: Dr. Fatemeh Keshavarzi, Ph.D, Assistant Professor, Department of Biology, Sanandaj Branch, Islamic Azad University, Sanandaj, Iran, Tel: +98 91 8370 4918; E-mail: gol.keshavarzi@gmail.com

Received Date: April 20, 2018; Accepted Date: October 26, 2018; Published Date: October 30, 2018

Citation: Arabzadeh F, Aeini F, Keshavarzi F, Behrvash S (2018) Resistance to Tetracycline and Vancomycin of *Staphylococcus aureus* Isolates from Sanandaj Patients by Molecular Genotyping. Ann Clin Lab Res Vol.6 No.4:260.

Abstract

Introduction: Studying antibiotic resistance of *Staphylococcus aureus* is very important and it has a main role in preventing creation of resistant strains. This study was done to determine the prevalence and genetic basis of tetracycline and vancomycin resistance in *Staphylococcus aureus* in Sanandaj.

Materials and methods: One hundred and fifty clinical isolates of *Staphylococcus aureus* were collected from Sanandaj Hospital. Susceptibility to antibiotics (tetracycline and vancomycin) were determined by disk agar diffusion method with minimal inhibitory concentration (MIC) evaluated on Muller-Hinton agar as described by the Clinical and Laboratory Standards Institute (CLSI). The tetracycline and vancomycin strains were screened by polymerase chain reaction (PCR) for the presence of five common vancomycin and tetracycline resistance determinants, respectively, van A, tet K, tet M, tet L and tet O.

Results: Using the DAD method, 12% of the *Staphylococcus aureus* isolates were resistant to vancomycin and 61/33% to tetracycline. For more, the tet (K) gene was found in 71 isolates, tet (L) in 5 isolates, tet (M) in 30 isolates and tet (O) were detected in one isolates and van A didn't see in any isolate by PCR technique.

Conclusion: This study indicates that resistance to tetracycline is mainly by efflux pumps mediated by tet (K) in *Staphylococcus aureus* in Sanandaj.

Keywords: *Staphylococcus aureus*; Antibiotic resistance; Tetracycline; Vancomycin; Sanandaj

Staphylococcus can grow in different environmental conditions but the best condition for their growth is 30 to 37°C. These bacteria are resistant and they can colonize on the skin and mucous membranes. *Staphylococcus aureus* species are one of the first known human pathogens. For the first time in 1880 and 1882 in laboratory and clinical studies, the disease caused by *staphylococcus* and its role in the growth of abscess was explained. Nowadays after 100 years, this organism is considered as one of the dangerous human pathogens. In general this bacterium is really important due to its tenacity destruction, potential power and increasing resistance to antibiotics [1]. *Staphylococcus* includes at least 40 species. Most of *staphylococcus* species are harmless and they reside on the skin and the mucosa of the human body and other animals. In the USA, *staphylococcus aureus* is responsible for the half of positive blood cultures among patients in ICU and it is one of the most important factors of pneumonia in these units [1,2]. The most population of *staphylococcus* is on the skin in human beings, especially in groin, armpit, perianal region and anterior nasal apertures and it can cause infection in all age groups. In terms of the place of getting infection, they can be divided into two groups: community acquired and hospital acquired. The content of cytosine +guanine in their DNAs is 30% to 40%. *Staphylococcus* species are anaerobic and they grow in the presence of bile salts and all of them are catalase-positive unlike *streptococci*. One of the prominent features in classifying the *staphylococcus* is producing coagulase enzyme [3]. Now 6 species of *staphylococcus* are coagulase-positive include *staphylococcus aureus*, *staphylococcus delphini*, *staphylococcus hyicus*, *staphylococcus intermedius*, *staphylococcus lutrae*, *staphylococcus pseudintermedius* and *staphylococcus schleiferi* the subspecies of coagulans. The most important species of coagulase-negative is *staphylococcus epidermiditis* which resides on the skin of human beings as a symbiotic (coexist). In the patients whose immune systems have been suppressed or those who use counters vessel, these bacteria can cause severe infections. *Staphylococcus saprophyticus* is another coagulase-negative species which is considered as a normal vaginal flora in women. This bacterium has a role in female genital tract infections among sexually active young women [4,5]. According to what was said, the purpose of this research is to

Introduction

During the last decade the prevalence of *staphylococcus aureus* species have been increasing worldwide.

study the prevalence and genetic basis of the resistance to antibiotics such as tetracycline and vancomycin in *staphylococcus aureus* samples in Sanandaj. The findings of this research may be helpful to choose the medicine and for better treatment in the region.

Materials and Methods

Analysis method

Samples: 150 *staphylococcus aureus* bacterial strains are collected from medical diagnostic laboratories in Sanandaj and Gram stain was done on all the samples to confirm that the bacterium is positive. After confirming Gram-positive bacteria, stock was provided from all collected samples which were on the Mueller-Hinton and blood agar. The biochemical differentiating tests to confirm the *staphylococcus aureus* included catalase, coagulase, DNAase, novobiocin sensitivity test and mannitol salt agar.

McFarland turbidity and antibiogram test

BaSo₄ was used according to the McFarland standard to standardize the inoculation concentration for the antimicrobial susceptibility test. The McFarland standard of BaSo₄ was prepared as following: half ml of (BaCl₂) 0.048 mol/l (W/V BaCl₂. 2H₂O% 1/175) was added to 99/5 ml sulfuric acid 0.18 mol/l (%1 V/V) and the suspension was prepared by constant stirring. The correct density of standard turbidity was determined through measuring the absorption in a spectrophotometer at optical path length of 1 cm then the

amount of 4-6 ml was poured into the screw-cap tubes, the same size as bacterial suspension tubes. After isolating the bacteria, some colonies of bacteria were removed by once (not loop) and they were dissolved in physiology serum. After preparing the homogeneous solution, it is stirred with a sterile swab then after rinsing off, the swab is transferred to the Mueller-Hinton medium and it is cultivated as meadow. Based on the FAO's table, if the bright halo of *staphylococcus aureus* is 14 mm or less, it is considered as a resistant strain and if this halo is 15 to 18 mm, it is considered as intermediate resistant strain and if the mentioned halo is 19 mm or more, the strain is considered as susceptible. For vancomycin, if the halo is 11 mm or less, the strain is resistant, if this halo is 11 to 14 mm, the strain is intermediate resistant and if the halo is 15 mm or more, it is susceptible.

DNA extraction and PCR

Genome of those colonies which were cultivated in the selective medium of cultures and those which biochemical tests were done on them and were determined as the *staphylococcus aureus* bacteria, were extracted by DNA extraction kit of Gram-positive bacteria (CinnaGen company). Then PCR was done by the primers of **Table 1**. Target genes in thermocycler were done through a plan consists of the initial denaturation in 94°C for 5 minutes and then 30 cycles of denaturation in 92°C for 30 seconds, annealing in 50 to 60°C for 30 seconds and extension in 72°C for 1 minute and then the final extension in 72°C for 10 minutes. Size marker of 100 (bp) was used to determine the size of bands in this research.

Table 1 Genes responsible for resistance and their primer's sequence.

Genes	Primer's Sequence of Genes
Tet (M)-R	5-TCCGACTATTTAGACGACGG-3
Tet (M)-F	5-AGTTTTAGCTCATGTTGATG-3
Tet (K)-R	5-GTAGTGACAATAAACCTCCTA-3
Tet (K)-F	5-GTAGCGACAATAGGTAATAGT-3
Tet (O)-R	5-TCCCACTGTTCCATATCGTCA-3
Tet (O)-F	5-AACTTAGGCATTCTGGCTCAC-3
Tet (L)-R	5-AACCAGCCAATAATGACAAGAT-3
Tet (L)-F	5-ATAAATTGTTTCGGGTCGGTAAT
Van (A)-R	5-TCACCCCTTTAACGCTAATA-3
Van (A)-F	5-ATGAATAGAATAAAAGTTGC-3

Results

In this research, first 192 samples of bacteria were collected then 150 samples of collected bacteria were used as *staphylococcus aureus* through biochemical and differentiation tests.

Antibiotic susceptibility test results

12% of *staphylococcus aureus* were resistant to vancomycin (18 out of 150 samples), 52% of samples were intermediate resistant (78 out of 150 samples) and 36% of them were susceptible to vancomycin (54 out of 150 samples) (**Figure 1**). In addition, according to the antibiotics test results, 61/33% of *staphylococcus aureus* were resistant to tetracycline (92 out of

150 samples), 26% were intermediate resistant (39 out of 150 samples) and 12/667% of them were susceptible to tetracycline (19 out of 150 samples) (Figure 2).

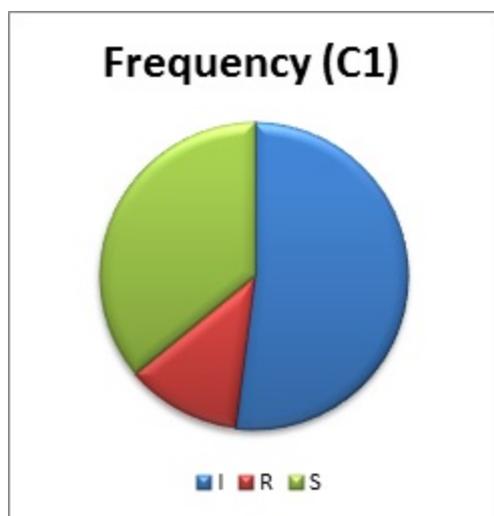


Figure 1 Relative frequency of resistant (R), susceptible (S) and intermediate resistant *Staphylococcus* to vancomycin (C1), 12% of *Staphylococcus aureus* samples (18 out of 150 samples) were resistant, 52% (78 out of 150 samples) were intermediate resistant and 36% (54 out of 150 samples) were susceptible to vancomycin.

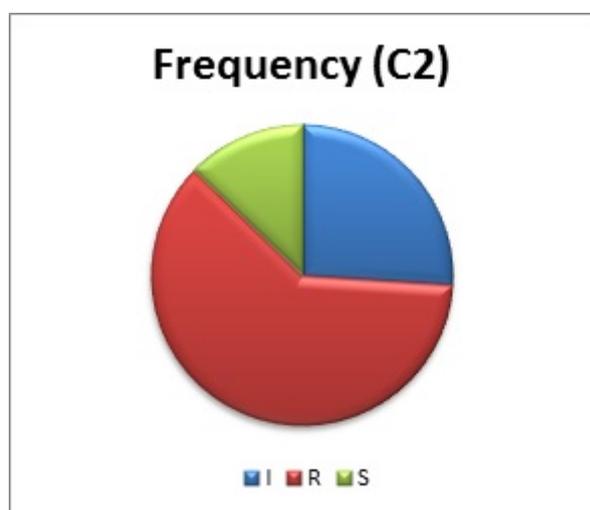


Figure 2 Pie chart of relative frequency of resistant (R), susceptible (S) and intermediate resistant *Staphylococcus* to tetracycline (C2), according to antibiotics test results, 61/33% of *Staphylococcus aureus* samples (92 out of 150 samples) were resistant, 26% (39 out of 150 samples) were intermediate resistant and 12/667% (19 out of 150 samples) were susceptible to tetracycline.

DNA extraction and PCR results of *Staphylococcus aureus*

According to the PCR results, 47/33% had tet (K) gene (71 out of 150 samples of *Staphylococcus aureus*). Also 60/86% had tet (K) gene (56 out of 92 samples of resistant *Staphylococcus aureus* to tetracycline) and it was observed in 15 samples of intermediate resistant samples (38/46% of intermediate resistant bacteria). Van (A) gene was not observed in this research. Also 21/33% had tet (M) gene (32 out of 150 samples of *Staphylococcus aureus*). Also 30% had tet (M) gene (28 out of 92 samples of tetracycline resistant *Staphylococcus aureus*). Four samples of tetracycline intermediate resistant *Staphylococcus aureus* had this gene. In addition, 21 out of 32 tet (M) *Staphylococcus aureus* samples had tet (K) gene as well (65/62%). Foremore 0/66% (1 out of 150 *Staphylococcus aureus* samples) and 1/8% of tetracycline resistant *Staphylococcus aureus* had tet (O) gene (1 *Staphylococcus aureus* bacterium of 92 bacteria). This sample had tet (K) gene as well. Results showed that 3/33% (5 out of 150 *Staphylococcus aureus* samples) and 5/43% of tetracycline resistant *Staphylococcus aureus* samples had tet (L) gene (5 *Staphylococcus aureus* bacteria of 92 bacteria). It should be mentioned that 3 samples of 5 *Staphylococcus* samples had both tet (L) and tet (K) gene (60% of tet (L) samples) and one sample of these 5 samples had tet (M) (20%). Only one sample out of 150 *Staphylococcus aureus* samples had three genes of tet (K), tet (M) and tet (L) (0/66%).

Discussion

According to National Nosocomial Infection Surveillance Committee, 75% of negative-coagulase *Staphylococcus* and 47% of positive-coagulase strains which were isolated from intensive care unit in December of 2000 were resistant to methicillin, so vancomycin is the first selective drug in the treatment of infections [6]. Vancomycin is widely used in the treatment of infections caused by methicillin resistant *Staphylococcus aureus* which leads to the increasing vancomycin resistant and the spread of species of heterogeneous resistant, intermediate and resistant bacteria in different parts of the world and it has created a great concern in clinical samples so that the number of the species of these vancomycin resistant bacteria are increasing.

Being vancomycin resistant, these bacteria are usually become resistant to a wide range of antibiotics. VISA (Vancomycin Intermediate resistance *Staphylococcus aureus*) and VRSA species have the potential for development and they are also increasing in our country. Vancomycin resistant *Staphylococcus aureus* is at the head of research topics because the increasing use of surgeries, dialysis and modern medical methods lead to the increasing use of vancomycin. Nowadays there are few drugs like vancomycin which cover this bacterium. Due to the above mentioned reason, this research was done in this regard. It is obvious that studying multi-drug resistance like simultaneous resistance to vancomycin and tetracycline, pave the way to detect resistant species more effectively. It is accepted as a general rule that

tetracycline resistance causes resistance to other antibiotics [7].

In this research, van (A) gene was not observed among 150 *staphylococcus aureus* samples. This result can be satisfying, for being vancomycin resistant to is really worrying and dangerous. At the moment, vancomycin is one of the few antibiotics which are effective in the treatment of *staphylococcus aureus* infections. The observed vancomycin resistance of this bacterium through disk diffusion method can be the result of phenotypic causes or environmental and human error. In addition the results can indicate that disk diffusion method is not accurate enough for this research. This result is consistent with the findings of some researchers in the field. Hong Binkim et al. (1999-2001) studied 682 *staphylococcus aureus* samples in Korea and no resistant or intermediate resistant sample was observed [8,9]. In a research which was done in Be sat hospital in Sanandaj, there was no vancomycin resistant strain or VRSA [10]. In another research, 5/6% was vancomycin resistant enterococcus but there was no VISA or VRSA [11]. In this research, 94/87% of vancomycin intermediate resistant *staphylococcus aureus* samples (74 out of 78) are resistant to methicillin (through disk diffusion). Also 94/44% of vancomycin resistant *staphylococcus aureus* samples (17 out of 18) are considered as MRSA samples through disk diffusion method, because most of vancomycin resistant, intermediate resistant are resistant to methicillin as well. Lack of resistant gene expression in all resistant samples to that gene, can be due to the different reasons, including phenotypic method or environmental and human error. In addition, observing phenotypic resistance is not a definite reason for resistant gene expression. In other words, lack of resistant gene in some antibiotic resistant samples could be also due to the lack of gene expression. For example, in studying van (A) gene, 12% of *staphylococcus samples* were identified as resistant ones through phenotypic method. In PCR stage, there was no resistant gene so it can be applicable to the above mentioned content. Huys et al. reported 58% of tet (K) gene in collected *staphylococcus aureus* from poultry slaughterhouses in South Africa [12]. In this research the prevalence of tetracycline resistance due to tet (K) gene was higher than other genes and this result is consistent with other researcher's findings in this field. The lowest resistance to tetracycline was related to tet (O) gene which is consistent with other researchers' findings too. As mentioned before, those *staphylococcus aureus* strains which only have tet (K) gene are tetracycline resistant, but they are susceptible to minocycline. Those strains which have tet (M) gene are resistant to all antibiotics in doxycycline group. These strains have tet (K) gene as well. Tet (L) gene is observed in the strains which have tet (M) gene. There was no report of tet (O) gene in *staphylococcus aureus* strains until 2004. *Staphylococcus aureus* has a wide distribution of tet (K) and tet (M) genes. These two genes have been located on mobile genetic elements such as small plasmids and transposons. It should be mentioned that this small plasmid is not able to transfer spontaneously but it can get the ability of movement from plasmids having conjugative genetic transmission power [13]. In addition, in this research 21 out of 32 tet (M) *staphylococcus*

aureus samples (65/62%) have tet (K) gene as well. According to Greet Huys [12], *staphylococcus aureus* has a wide distribution of tet (K) and tet (M) genes which is in consistent with the result of this research. Also a tet (O) *staphylococcus aureus* strain has tet (K) as well. 3 out of 5 samples of tet (L) *staphylococcus aureus* have tet (K) too (60%). Only 1 out of 5 samples of tet (L) *staphylococcus aureus*, has tet (M) gene (20%). Among 150 *staphylococcus aureus* samples, only one sample had three tet (K), tet (M) and tet (L) genes (66% of all *staphylococcus aureus* samples). In other words, these three genes were expressed just in one sample.

Conclusion

This study indicates that resistance to tetracycline is mainly by efflux pumps mediated by tet (K) in *Staphylococcus aureus* in Sanandaj.

Author's Contributions

Fatemeh keshavarzi designed the study, Fatemeh keshavarzi and Fatemeh Aeini wrote the paper; Foozieh Arabzadeh and Pezhman Karami, conceived the experiments, prepared the Figures; collected the samples. All authors gave final approval for the manuscript to be submitted for publication.

Acknowledgments

This work was supported by the Islamic Azad University, Sanandaj Branch. Research is based on Miss. Foozieh Arabzadeh's master's thesis. The authors would like to also extend their sincere appreciation to School of Medicine, Kurdistan University of Medical Sciences, Sanandaj, Iran.

Conflict of Interest

The authors declare no conflict of interests.

References

1. Franklin D, Lowy MD (1998) *Staphylococcus aureus* Infection. N Engl J Med 339: 520- 532.
2. Hauschild T, Kehrenberg C, Schwarz CT (2003) Etracycline resistance in *staphylococci* from free-living rodents and insectivores. J Vet Med Series B Infect Dis Vet Publ Health 50: 443-446.
3. Akoua Koffi C, Dje K, Toure R, Guessennd N, Acho B, et al. (2004) Nasal carriage of methicillin-resistant *Staphylococcus aureus* among health care personnel in Abidjan. Dakar Med 49(1): 70-74.
4. Chopra I, Roberts M (2001) Tetracycline antibiotics: Mode of action, applications, molecular biology, and epidemiology of bacterial resistance. Microbiol Mol Biol Rev 65(2): 232-260.
5. Klein E, Smith DL, Laxinarayan R (2007) Hospitalizations and deaths caused by methicillin-resistant *Staphylococcus aurus*, United States, 1999-2005. Emerg Infect Dis 13(12): 1840-1860.
6. Jones CH, Tuckman M, Howe AY, Orłowski M, Mullen S, et al. (2006) Diagnostic PCR analysis of the occurrence of methicillin

- and tetracycline resistance genes among *Staphylococcus aureus* isolates from phase 3 clinical trials of tigecycline for complicated skin and skin structure infections. *Antimicrob Agents Chemother* 50(2): 505-510.
7. Locksley RM (2005) *Staphylococcal* infections in Harrison's principles of internal medicine, 16th Edition, volum I, United States of America, Mc Graw-Hill companies pp: 745-768.
 8. Kim HB, Park WB, Lee KD, Choi YJ, Park SW, et al. (2003) National Surveillance for *Staphylococcus aureus* with reduced susceptibility to Vancomycin in Korea. *J Clin Microbiol* 41(6): 2279-2281.
 9. Wang G, Hindler GF, Ward KW, Brunckner DA (2006) Increased vancomycin MICs for *Staphylococcus aureus* clinical isolates from a university hospital during a 5 year period. *J Clin Microbiol* 44(11): 3883-3886.
 10. Rashidian M, Taherpoor A, Goodarzi S (2001) Nasal carrier rates and antibiotic resistance of *staphylococcus aureus* isolates of Beasat hospital staff. *Scientific Journal of Kurdistan University of Medical Sciences* 6(1): 1-8.
 11. Ghasemian R, Najafia N, Shojai A (2004) Nasal carriage and antibiotic resistance of *staphylococcus aureus* isolates of Razi hospital personel, Qaemshahr. *J Mazandaran Univ Med Sci* 14(44): 79-86.
 12. Huys G, Cnockaert M, Vaneechoutte M, Woodford N, Nemec A, et al. (2005) Distribution of tetracycline resistance genes in genotypically related and unrelated multiresistant *Acinetobacter baumannii* strains from different European hospitals. *Res Microbiol* 156(3): 348-355.
 13. Tyagi S, Oberoi A (2015) Prevalence of inducible clindamycin resistance among *Staphylococcal* isolates in a tertiary care hospital in North India. *Indian J Med Microbiol* 33: 327-328.